# Current State of Insecticide Resistance in (Florida) Mosquitoes

#### Dodd 2024- History of Insecticide Resistance

Tuesday, January 30, 2024 Module 2

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USDA ARS Center for Medical Agricultural & Veterinary Entomology
Mosquito & Fly Research Unit



# Current State of Insecticide Resistance in (Florida) Mosquitoes

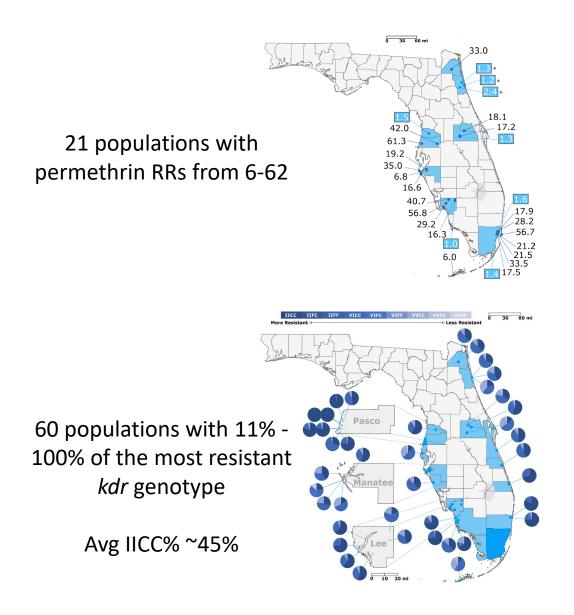
- 1. IR today: What we know, what we don't
- 2. IR mechanisms
- 3. Why mechanism matters:

  Aedes aegypti vs. Culex quinquefasciatus
- 4. Future trends in IR



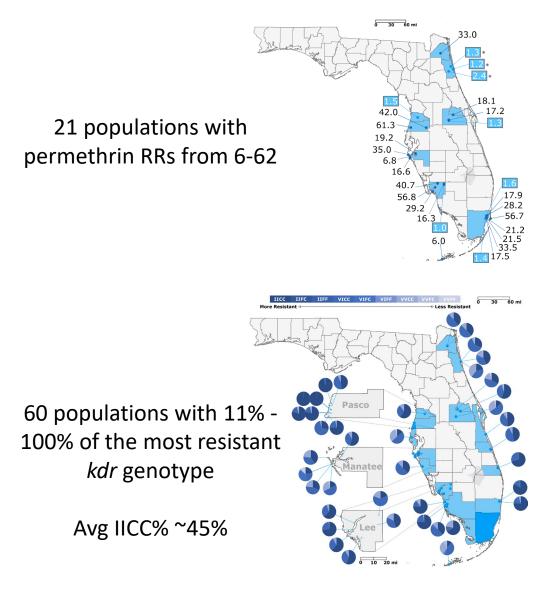


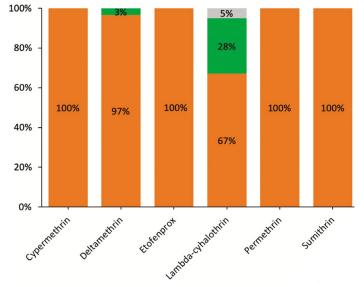
# Most populations of *Aedes aegypti* are resistant to pyrethroids



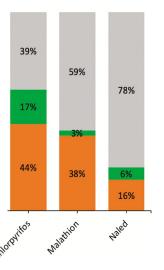


# Most populations of Aedes aegypti are resistant to pyrethroids





33 populations 100% resistant to permethrin 97% resistant to deltamethrin



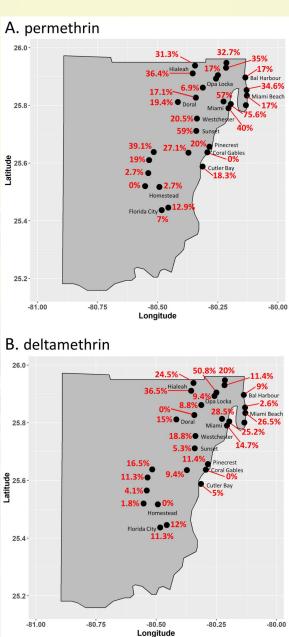
Resistance to OPs was variable and tended to be relatively weak

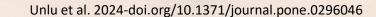


### Insecticide Resistance in Miami Culex quinquefasciatus

All Miami populations resistant to permethrin

All Miami populations resistant to deltamethrin

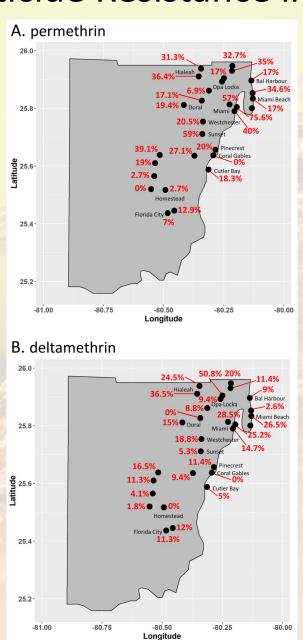


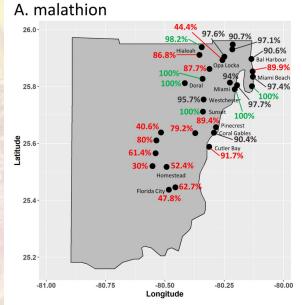


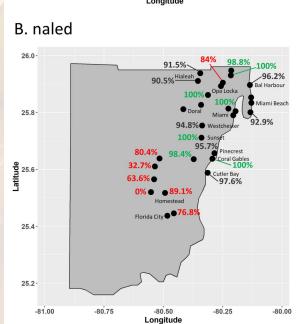
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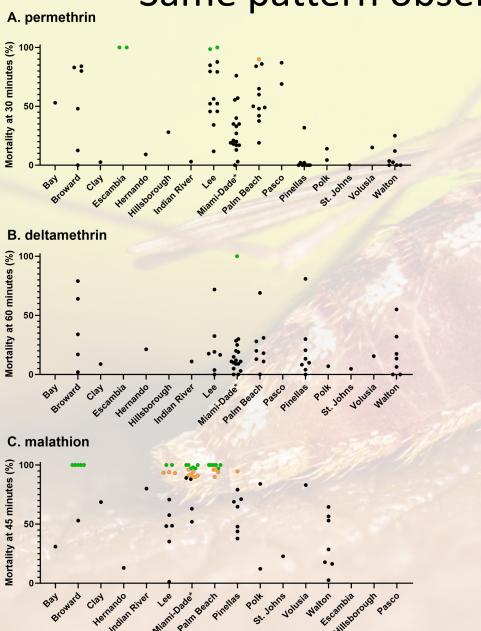




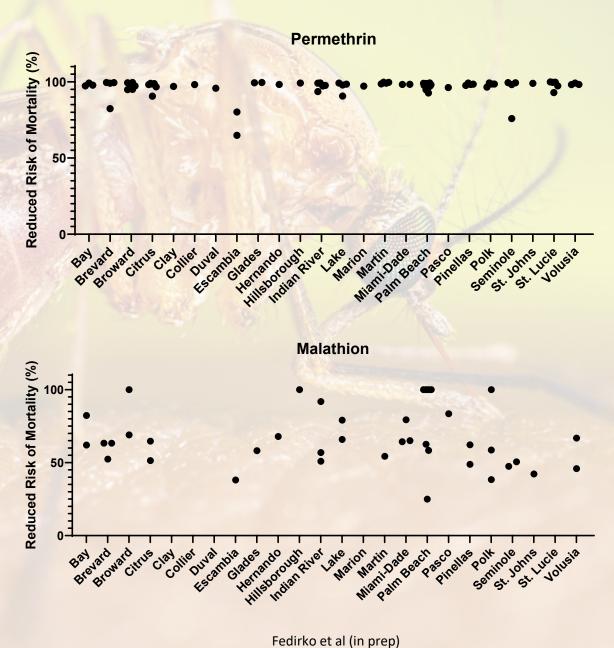
Mixture of IR intensity to malathion

IR can be strong enough to reduce Naled efficacy

# Same pattern observed across the state of Florida



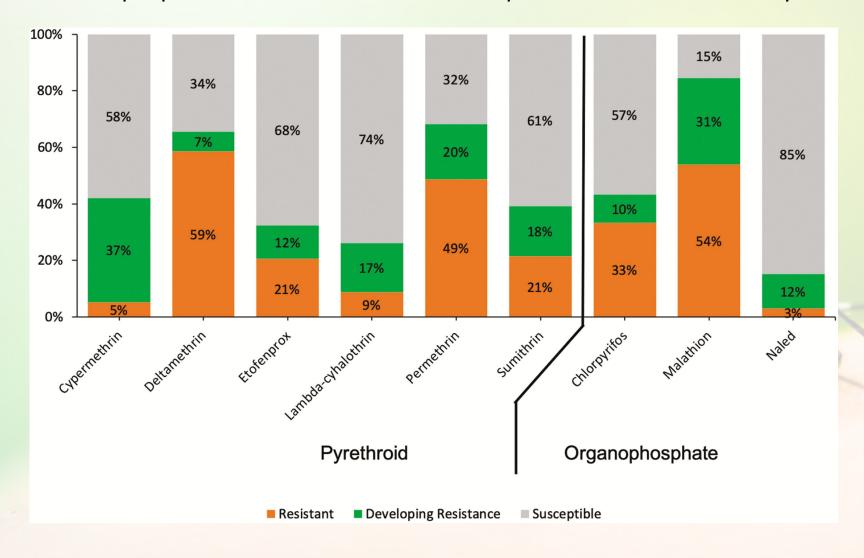
Estep et al (in prep)



Most populations of Aedes albopictus are relatively susceptible to adulticides

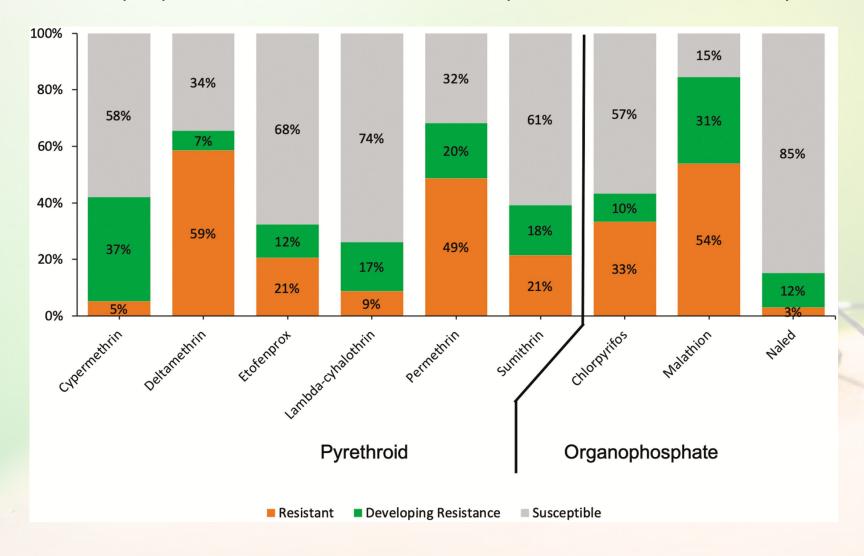


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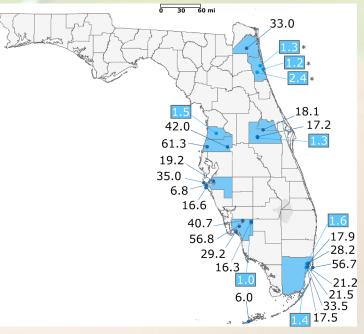
Low level IR to permethrin, deltamethrin, malathion & Naled in some populations

#### Most populations of Aedes albopictus are relatively susceptible to adulticides



Low level IR to permethrin, deltamethrin, malathion & Naled in some populations

Topical application confirms that the IR to pyrethroids is weak (RR<3)



IR Status of other species

Cx. nigripalpus

Ae. taenforkynchus

An. crucians

An. quadrimaculatus



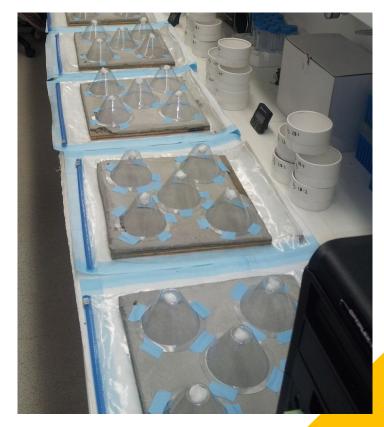
- Behavioral resistance
- Penetration (cuticular) resistance
- Metabolic resistance
- Target-site resistance

- Behavioral resistance
  - Alter mobility behavior to avoid contamination
  - Alter reproductive behaviors
  - Fire ants
    - Remove unhealthy materials
    - Grooming of self and others
    - Cover toxic areas

"...behavior that reduces an insect's exposure to toxic compounds or that allows an insect to survive in what would otherwise be a toxic and fatal environment."

Sparks, T. C., Lockwood, J. A., Byford, R. L., Graves, J. B., and Leonard, B. R. (1989). The role of behavior in insecticide resistance. Pestic. Sci. 26, 383–399. doi:10.1002/ps.2780260406

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Dunford, J.C., Estep, A.S., Waits, C.M., Richardson, A.G., Hoel, D.F., Horn, K., Walker, T.W., Blersch, J.S., Kerce, J.D. and Wirtz, R.A., 2018. Evaluation of the long-term efficacy of K-Othrine® PolyZone on three surfaces against laboratory reared Anopheles gambiae in semi-field conditions. *Malaria Journal*, *17*(1), pp.1-7.

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Photo Credit: James Gathany Content Providers(s): CDC

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https://www.pctonline.com/news/seven-tips-blow-fly-management/

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- Penetration (cuticular) resistance
  - Resistant An. gambiae have thicker cuticles
  - Silencing a cuticle gene in resistant Cx. pipiens (quinqs) = decreased cuticle

"Insects create barriers...using their outer cuticle, which protects them against a wide spectrum of insecticides."

Siddiqui JA, Fan R, Naz H, Bamisile BS, Hafeez M, Ghani MI, Wei Y, Xu Y and Chen X (2023), Insights into insecticide resistance mechanisms in invasive species: Challenges and control strategies. Front. Physiol. 13:1112278.

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- Metabolic resistance
  - Resists multiples classes of Als
  - Multiple classes of enzymes
    - Carboxylesterases
    - Glutathione S-transferases
    - Cytochrome P450
  - Synergists help with
    - Mosquito control
    - Test for metabolic resistance

"Metabolic resistance is a type of resistance inferred by metabolic activities in insects that help them detoxify or break down contaminants or the ability to eliminate toxic compounds from their bodies more quickly."

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|                  |                 | CMAVE | LL    | I F   | FF    |
|------------------|-----------------|-------|-------|-------|-------|
| Permethrin       | LD50            | 1.08  | 15.12 | 21.29 | 29.46 |
| Permethrin + PBO | LD50            | 0.12  | 0.61  | 1.12  | 2.01  |
|                  | Synergist ratio | 9.00  | 24.79 | 19.01 | 14.66 |

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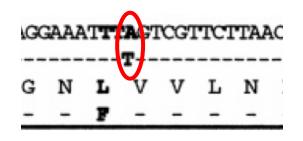
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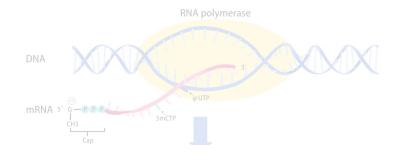
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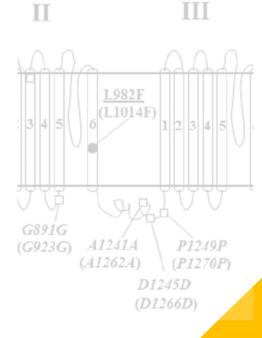
https://lite.lstmed.ac.uk/lite-facilities/lite-laboratories/cdc-bottle-assay

• Target-site resistance



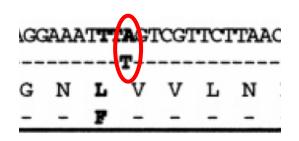


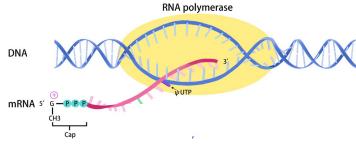
DNA RNA

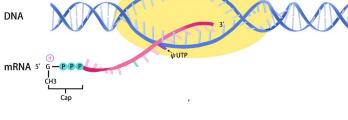


protein

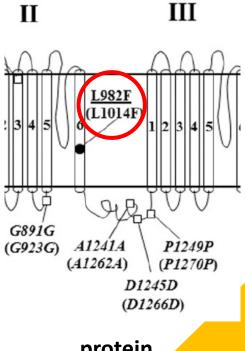
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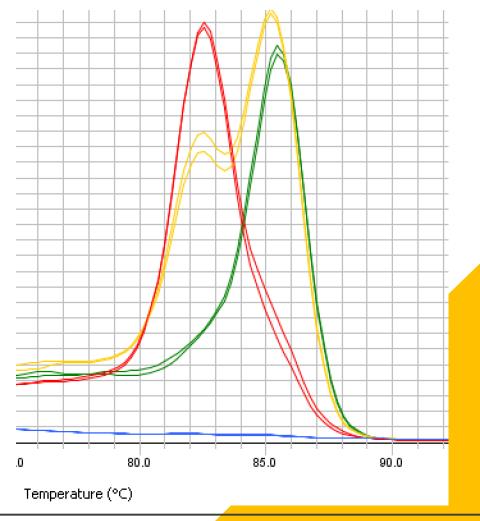
DNA **RNA** 



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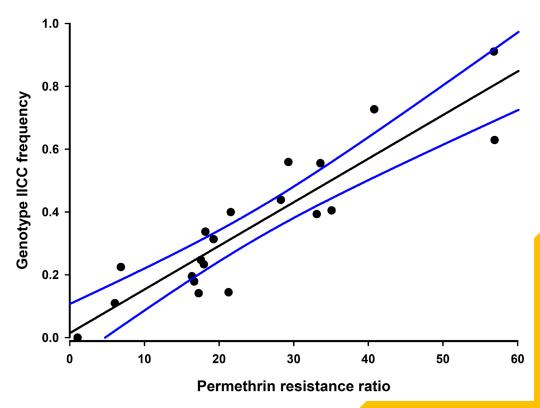
## IR mechanisms underlying the resistance

- Target-site resistance
  - *kdr* knock-down resistance
    - NaV gene
    - *Ae. aegypti: kdr* = PYR resistance
  - Many SNPs



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Estep et al. 2018. Quantification of permethrin resistance and kdr alleles in Florida strains of Aedes aegypti (L.) and Aedes albopictus (Skuse). *PLoS neglected tropical diseases*, *12*(10), p.e0006544.

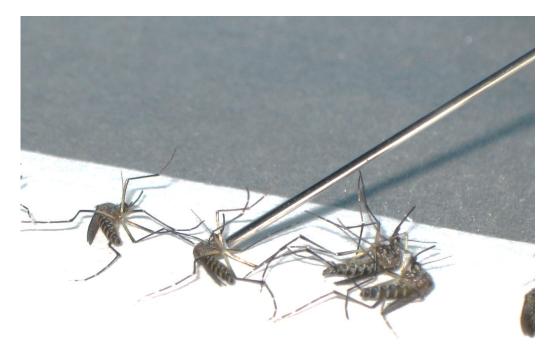
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| Mutation            | Strain              | n* | Phenotype <sup>†</sup> | Co                           | odons‡ (Frequency [%] ± SE)  |                             |
|---------------------|---------------------|----|------------------------|------------------------------|------------------------------|-----------------------------|
| A109S <sup>5</sup>  | S-Lab               | 60 | Susceptible            | GCA (65±5.0)                 | G/TCA (35±5.0)               | TCA (0)                     |
|                     | MAmCq GO            | 60 | 10-fold                | GCA (0)                      | <u>G/T</u> CA (0)            | TCA (100)                   |
|                     | MAmCq G6            | 60 | 570-fold               | GCA (0)                      | G/TCA (0)                    | TCA (100)                   |
| L982F1              | S-Lab               | 60 | Susceptible            | TTA (100)                    | TTA/T (0)                    | TTT (0)                     |
|                     | MAmCq <sup>G0</sup> | 60 | 10-fold                | $TT\overline{A}$ (22 ± 3.0)  | TTA/T (52 ±6.0)              | TT <u>T</u> (26 ±7.5)       |
|                     | MAmCq <sup>G6</sup> | 60 | 570-fold               | TT <u>A</u> (0)              | TT <u>A/T</u> (0)            | TTT (100)                   |
| W1573R <sup>a</sup> | S-Lab               | 60 | Susceptible            | TGG (100)                    | T/C <del>GG</del> (0)        | CGG (0)                     |
|                     | MAmCq <sup>G0</sup> | 60 | 10-fold                | $\overline{1}$ GG (72± 10.5) | T/CGG (25 ±8.5)              | $\overline{C}GG$ (3.0 ±3.0) |
|                     | MAmCq <sup>G6</sup> | 60 | 570-fold               | TGG (0)                      | T/CGG (8 ±5.5)               | CGG (92 ±6.0)               |
| L852L#              | S-Lab '             | 60 | Susceptible            | CTG (100)                    | CTG/A (0)                    | CTA (0)                     |
|                     | MAmCq GO            | 60 | 10-fold resistance     | CTG (27±10)                  | $CT\overline{G/A}$ (38±7.5)  | $CT\overline{A}$ (35±5)     |
|                     | MAmCq G6            | 60 | 570-fold resistance    | СТ <del>С</del> (0)          | $CTG/A (6.5 \pm 2.8)$        | $CT\overline{A}$ (93.5±2.9) |
| G891G#              | S-Lab .             | 60 | Susceptible            | GGC (100)                    | GGC/A (0)                    | GGĀ (0)                     |
|                     | MAmCq GO            | 60 | 10-fold resistance     | GGC (28±10)                  | GGC/A (42±7.5)               | $CT\overline{A}$ (30±10)    |
|                     | MAmCq G6            | 60 | 570-fold               | GG <u>C</u> (0)              | GG <del>C/A</del> (5±5)      | CTA (95±5)                  |
| A1241A#             | S-Lab .             | 60 | Susceptible            | GCA (100)                    | GCA/G (0)                    | $GC\overline{G}$ (0)        |
|                     | MAmCq GO            | 60 | 10-fold resistance     | $GC\overline{A}$ (2±3)       | GCA/G (18±2.9)               | GCG (80±5.5)                |
|                     | MAmCq G6            | 60 | 570-fold               | $GC\overline{A}(0)$          | GCA/G (0)                    | GCG (100)                   |
| D1245D#             | S-Lab '             | 60 | Susceptible            | GA <del>C</del> (100)        | GAC/T (0)                    | GAT (0)                     |
|                     | MAmCq GO            | 60 | 10-fold resistance     | GAC (38±7.5)                 | $GA\overline{C/T}$ (45±8.5)  | $GAT (17\pm5.5)$            |
|                     | MAmCq G6            | 60 | 570-fold               | GA <del>C</del> (0)          | GAC/T (8±5.5)                | GAT (92±5.5)                |
| P1249P#             | S-Lab .             | 60 | Susceptible            | CCG (100)                    | CCG/A (0)                    | CCĀ (0)                     |
|                     | MAmCq GO            | 60 | 10-fold resistance     | $CC\overline{G}$ (37±5.5)    | $CC\overline{G/A}$ (42±5.5)  | $CC\overline{A}$ (21±5.5)   |
|                     | MAmCq G6            | 60 | 570-fold               | $CC\overline{G}(0)$          | $CC\overline{G/A}$ (5.0±5.0) | $CC\overline{A}$ (95±5.0)   |
| G1733G#             | S-Lab '             | 60 | Susceptible            | $GG\overline{A}$ (48 ± 12.5) | GGA/G (52±12.5)              | GG <u>G</u> (0)             |
|                     | MAmCq GO            | 60 | 10-fold resistance     | GGĀ (0)                      | GGA/G (5±5.0)                | GGG (95±5.0)                |
|                     | MAmCq G6            | 60 | 570-fold resistance    | GG <u>A</u> (0)              | GG <u>A/G</u> (0)            | GGG (100)                   |

Li, T., Zhang, L., Reid, W.R., Xu, Q., Dong, K. and Liu, N., 2012. Multiple mutations and mutation combinations in the sodium channel of permethrin resistant mosquitoes, Culex quinquefasciatus. *Scientific Reports*, *2*(1), p.781.

Ae. aegypti



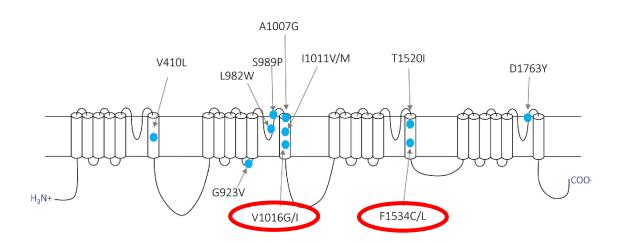
Cx. quinquefasciatus



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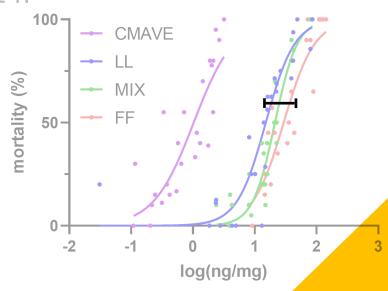
#### Ae. aegypti

• V1016I & F1534C



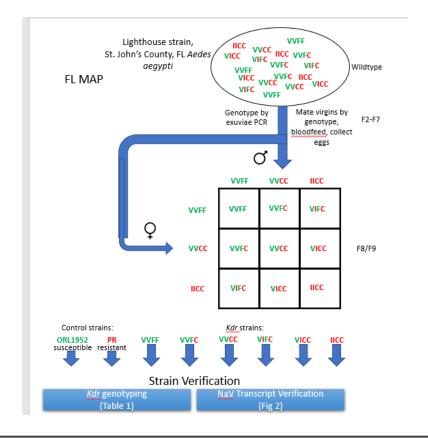
Chen, M., Du, Y., Nomura, Y., Zhorov, B.S. and Dong, K., 2020. Chronology of sodium channel mutations associated with pyrethroid resistance in Aedes aegypti. *Archives of insect biochemistry and physiology*, *104*(2), p.e21686.

#### Cx. quinquefasciatus

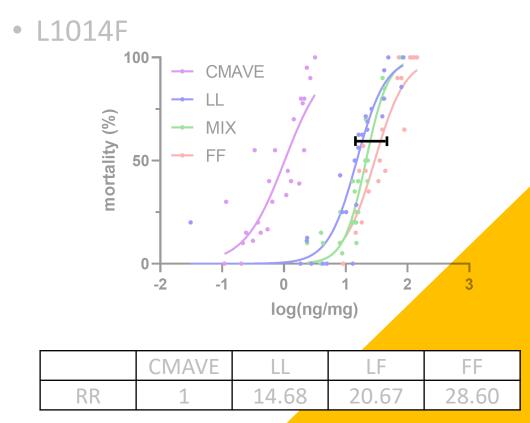


|    | CMAVE | LL    | LF    | FF    |
|----|-------|-------|-------|-------|
| RR | 1     | 14.68 | 20.67 | 28.60 |

Ae. aegypti

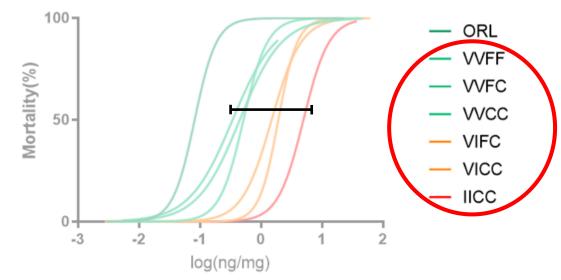


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Ae. aegypti

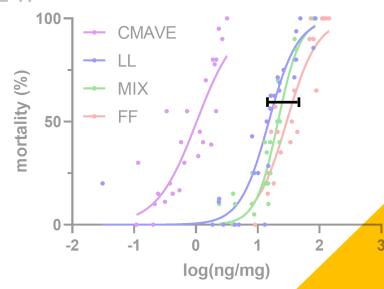
• V1016I & F1534C



|    | ORL | VVFF | VIFC | IICC |
|----|-----|------|------|------|
| RR | 1   | 5.5  | 23.5 | 60.7 |

Cx. quinquefasciatus

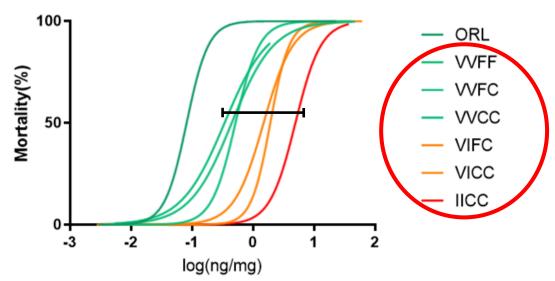




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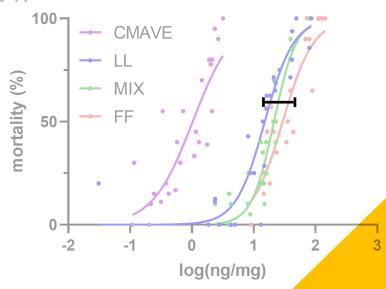
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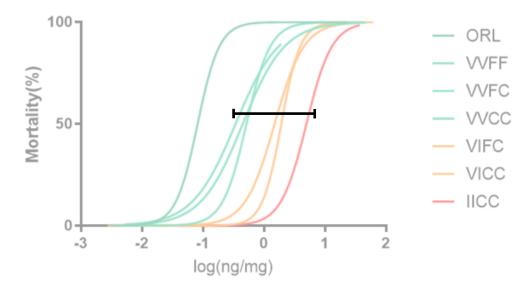
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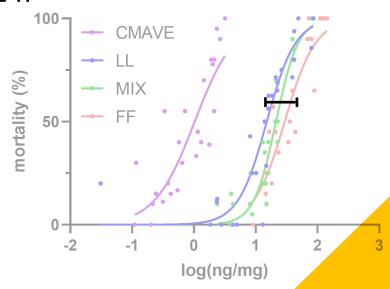


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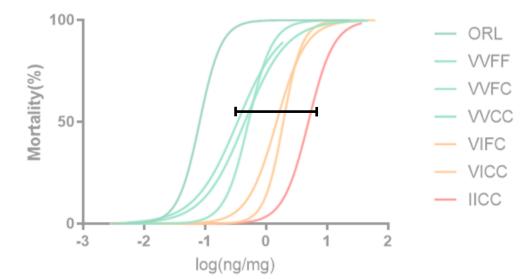
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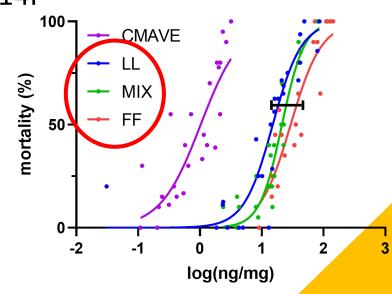


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|----|-----|------|------|------|
| RR | 1   | 5.5  | 23.5 | 60.7 |

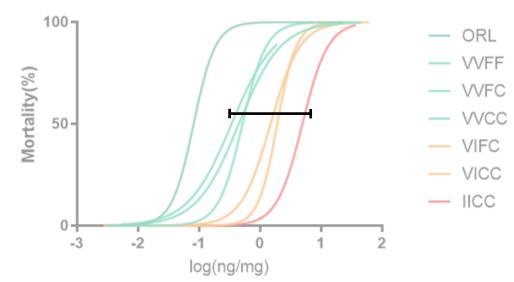
#### Cx. quinquefasciatus



|    | CMAVE | LL    | LF    | FF    |
|----|-------|-------|-------|-------|
| RR | 1     | 14.68 | 20.67 | 28.60 |

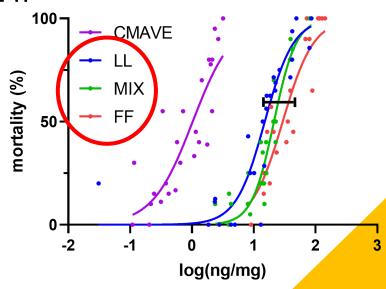


• V1016I & F1534C



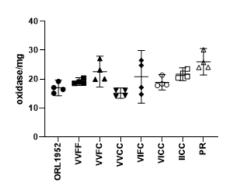
|    | ORL | VVFF | VIFC | IICC |
|----|-----|------|------|------|
| RR | 1   | 5.5  | 23.5 | 60.7 |

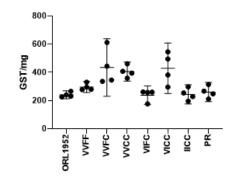
#### Cx. quinquefasciatus

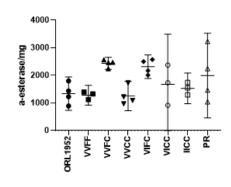


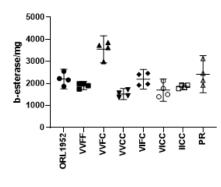
|    | CMAVE | LL    | LF    | FF    |
|----|-------|-------|-------|-------|
| RR | 1     | 14.68 | 20.67 | 28.60 |



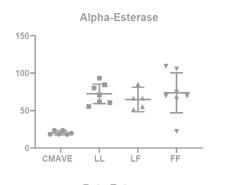


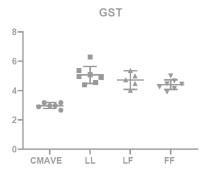


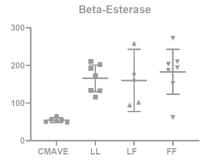


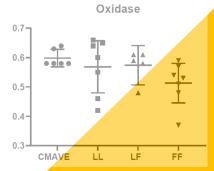


#### Cx. quinquefasciatus

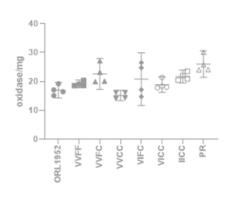


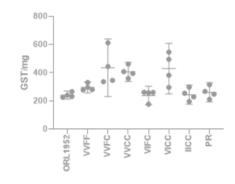


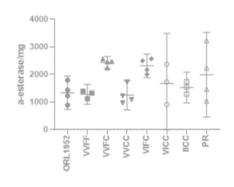


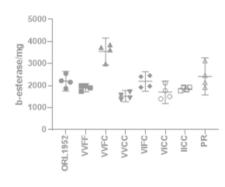


Ae. aegypti

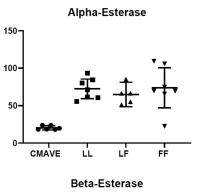


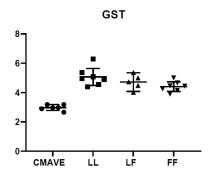


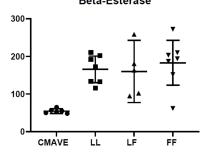


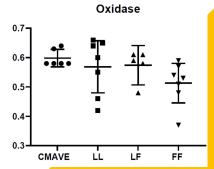


#### Cx. quinquefasciatus









Ae. aegypti

 Topically applied permethrin + PBO = low SR

|                  |                 | ORL   | VVFF | VIFC | IICC |
|------------------|-----------------|-------|------|------|------|
| Permethrin       | LD50            | 0.081 | 0.44 | 1.91 | 4.92 |
| Permethrin + PBO | LD50            | 0.12  | 0.37 | 1.61 | 3.28 |
|                  | Synergist ratio | 0.7   | 1.2  | 1.2  | 1.5  |

Cx. quinquefasciatus

|                  |                 | CMAVE | LL    | LF    | FF    |
|------------------|-----------------|-------|-------|-------|-------|
| Permethrin       | LD50            | 1.08  | 15.12 | 21.29 | 29.46 |
| Permethrin + PBO | LD50            | 0.12  | 0.61  | 1.12  | 2.01  |
|                  | Synergist ratio | 9.00  | 24.79 | 19.01 | 14.66 |

Ae. aegypti

 Topically applied permethrin + PBO = low SR

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Cx. quinquefasciatus

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#### Cx. quinquefasciatus

|                  |                 | CMAVE | LL    | LF    | FF    |
|------------------|-----------------|-------|-------|-------|-------|
| Permethrin       | LD50            | 1.08  | 15.12 | 21.29 | 29.46 |
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Ae. aegypti

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|                  | Synergist ratio | 0.7   | 1.2  | 1.2  | 1.5  |

Cx. quinquefasciatus

|                  |                 | <b>CMAVE</b> | LL    | LF    | FF    |
|------------------|-----------------|--------------|-------|-------|-------|
| Permethrin       | LD50            | 1.08         | 15.12 | 21.29 | 29.46 |
| Permethrin + PBO | LD50            | 0.12         | 0.61  | 1.12  | 2.01  |
|                  | Synergist ratio | 9.00         | 24.79 | 19.01 | 14.66 |

#### Ae. aegypti

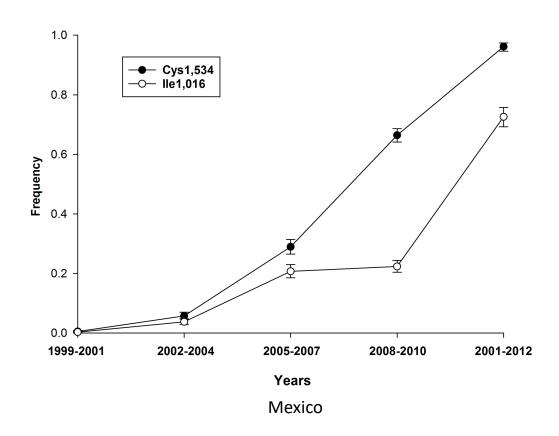
- Target site resistance = major mechanism
  - pyrethroid IR increases with # of IC alleles (>10X difference)
- Metabolic resistance ≠ major mechanism
  - No enzyme upregulation
  - Low SPs

#### Cx. quinquefasciatus

- Target site resistance ≠ major mechanism
  - kdr accounts for only <2X difference in pyrethroid IR
- Metabolic resistance = major mechanism
  - Increased enzyme upregulation
  - High SPs

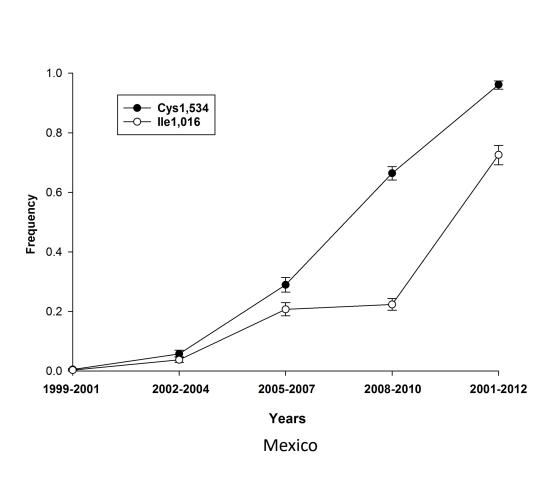
#### Examples indicate that IR will continue to build to pyrethroids

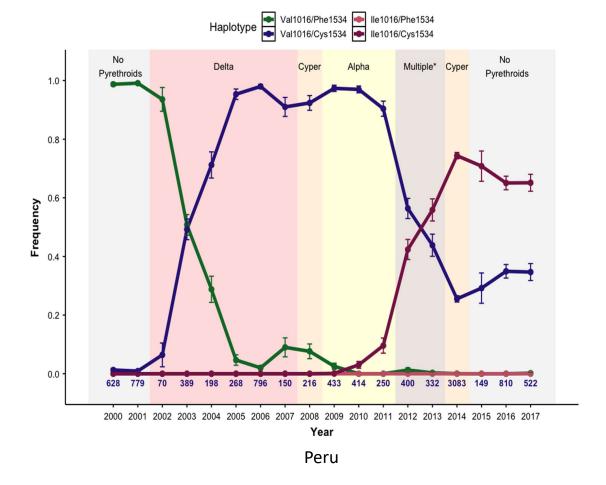
• The increase of the resistant genotypes is the standard in Aedes aegypti



Historical examples indicate that IR will continue to build to pyrethroids

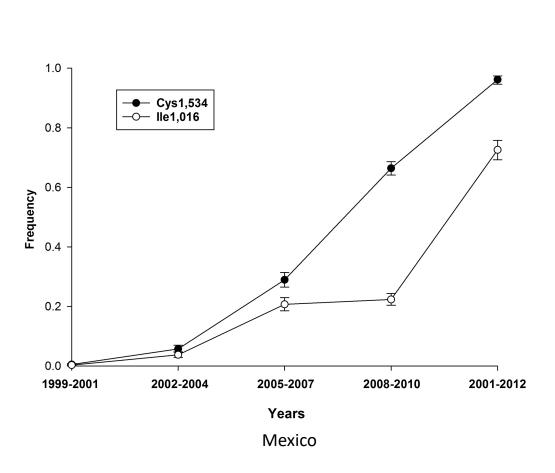
• The increase of the resistant genotypes is the standard in Aedes aegypti

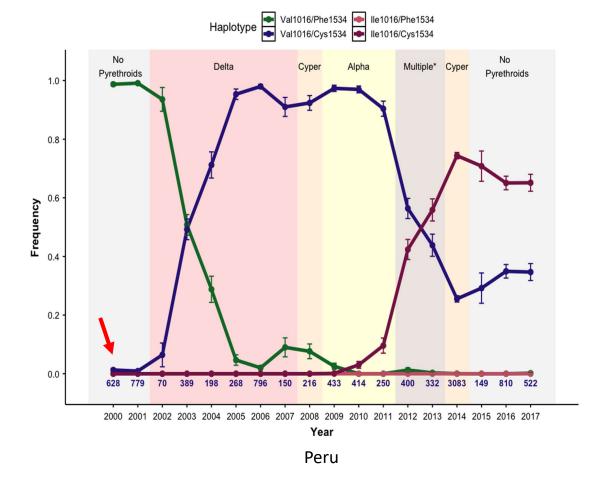




Historical examples indicate that IR will continue to build to pyrethroids

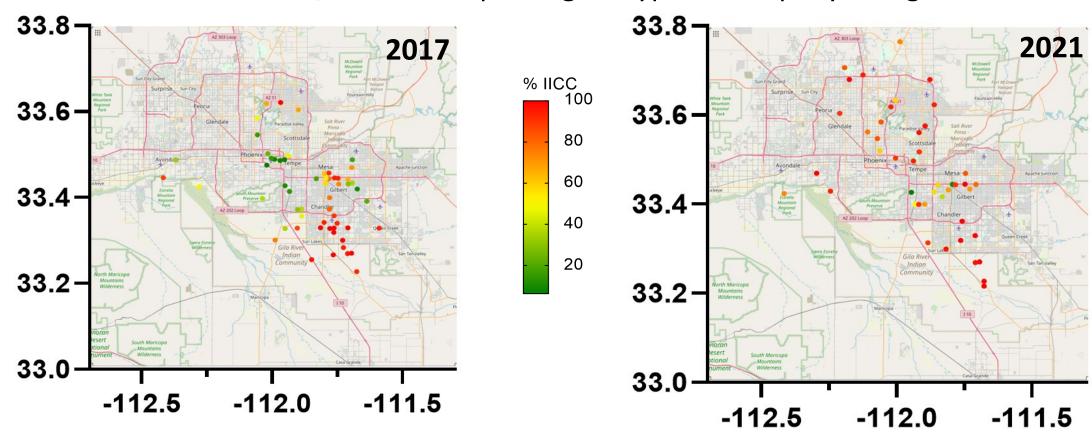
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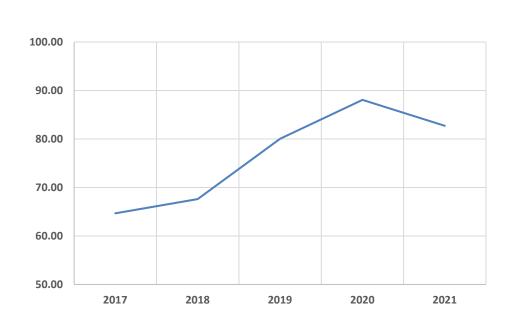
Historical examples indicate that IR will continue to build to pyrethroids

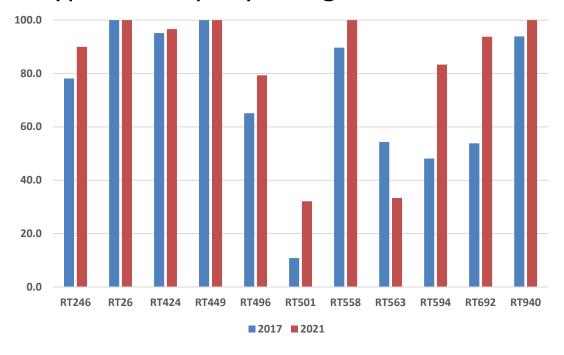
- The same appears to be occurring in US populations
  - Houston populations appear to be holding steady or increasing in IR
  - In Arizona, more susceptible genotypes are rapidly being lost



Historical examples indicate that IR will continue to build to pyrethroids

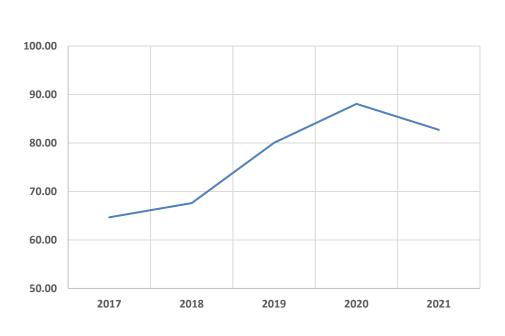
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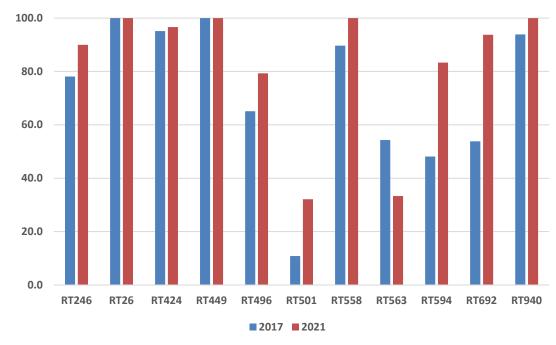




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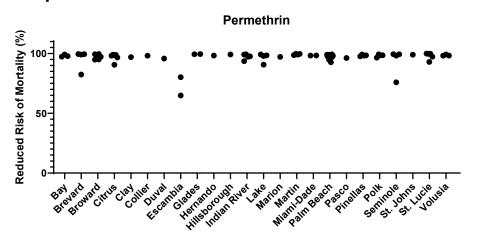


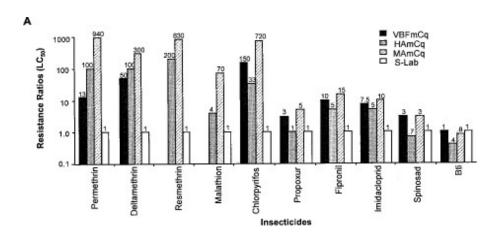


Currently, IR to OPs is low but there is history in Ae. aegypti

Historical examples indicate that IR will continue to build to pyrethroids & OPs

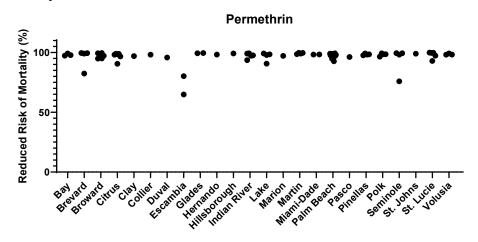
Culex quinquefasciatus are resistant but it can get much worse with more pressure



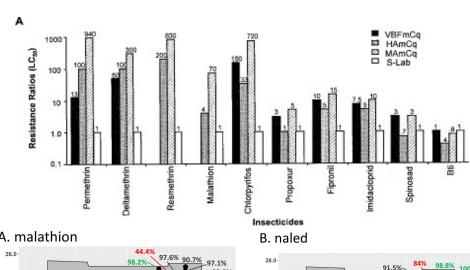


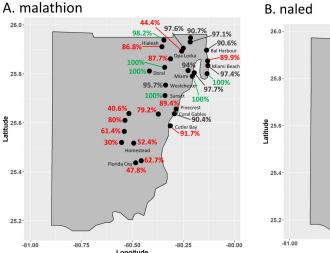
Historical examples indicate that IR will continue to build to pyrethroids & OPs

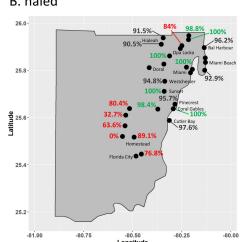
Culex quinquefasciatus are resistant but it can get much worse with more pressure



OPs may or may not be effective
 →Mechanism unknown







Unlu et al. 2024-doi.org/10.1371/journal.pone.0296046 Fedirko et al (in prep) Liu et al 2004-doi.org/10.1603/0022-2585-41.3.408

# Current State of Insecticide Resistance in (Florida) Mosquitoes

- 1. IR today: What we know, what we don't
- 2. IR mechanisms
- 3. Why mechanism matters: *Aedes aegypti* vs. *Culex quinquefasciatus*
- 4. Future trends in IR



# Thank you! Questions?

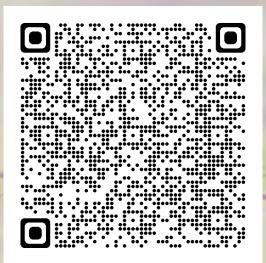
Just reach out...

<u>alden.estep@usda.gov</u> neil.sanscrainte@usda.gov

Alden Estep – Research Entomologist Neil Sanscrainte – Molecular Biologist







USDA ARS Center for Medical Agricultural & Veterinary Entomology

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