

# Variation When Determining dNDF and NDFD and Its Prediction by NIRS

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# Introduction

- **Nutritional importance of NDFD**
  - 2001 Dairy NRC suggests it can be used to determine dNDF for estimating TDN1X
  - Oba and Allen () indicates it is related to intake of lactating cows
- **Difference between NDFD and dNDF**
  - NDF Digestibility = NDFD (% of NDF) = digestion coefficient of NDF
  - digestible NDF = dNDF (% of DM) = proportion of DM that is digestible NDF
  - $dNDF = NDF * NDFD / 100$ 
    - 24% dNDF = 40% NDF \* 60% NDFD / 100
    - dNDF is always less than NDF

# Introduction

- **Methods of determining NDFD**
  - In vivo – using total collection or markers
    - Lactating cows fed mixed diets
    - Sheep at maintenance fed forage only
  - In situ – using porous bags
  - In vitro
    - Using flasks or tubes
    - Using filter bags – Ankom Daisy system
  - **Estimated using chemical composition**
    - Related to lignin and silica

# In Vivo Digestibility

- Is a biological evaluation of a feed that is not a constant, but varies with
  - Species
  - Size
  - Production level
  - Intake
  - Selection and sorting
  - Methodology

# Digestibility as a Measure of Animal Performance

- In vivo production digestibility protocol
  - Specific for the performance status of animals
  - Production level of intake (1-5X Mnt)
  - Ad libitum (free choice) intake with refusals = selection
  - Measures digestibility during production
  - Much greater variability = difficult to measure inputs and outputs

# Digestibility as a Measure of Feed Nutritive Value

- Standardized in vivo digestibility protocol
  - Designed to assign a value to a feed by minimizing animal performance differences
  - Mature animals
  - Maintenance level of intake (1X Mnt)
  - No selection or refusals
  - Measures maximum digestibility
  - Weigh feed, refusals and feces for 5-7 days

# In Situ / In Sacco Digestibility

- Feed is sealed in a porous bag and suspended in the rumen of fistulated cows
- Assume in situ = in vivo
  - But only measures fermentative digestion
- Apparent value is in mimicking ruminal digestion for production levels and diets
- More difficult to standardize, especially among labs when used for feed evaluation
  - Bag dimensions and pore sizes
  - Washing of bags and removal of fines
  - Cyclic and variable ruminal conditions
  - Variability among animals

# In Vitro Digestibility

- **Single-stage IVDMD**
  - Incubate ruminal fluid with feed in buffer
  - Dry residues and weigh
- **Two-stage Tilley & Terry IVDMD**
  - Incubate ruminal fluid with feed in buffer
  - Incubate undigested residue in acid pepsin
  - Dry residues and weigh

# In Vitro Digestibility

## Two-stage Van Soest IVDMTD

- Incubate ruminal fluid with feed in buffer
- Extract undigested residue in neutral detergent
- Dry NDF residues and weigh
- **In vitro methods measure different things**
  - Single and two-stage T&T IV measure apparent DM digestibility
  - Two-stage Van Soest IV measures true DM digestibility
  - T&T IVDMD will always be lower than VS IVDMTD

# Digestibility is a Variable

- NDFD and dNDF are a function of the feed and system in which it is measured
  - Not simply a feed characteristic
  - In vivo digestibility is affected by the animal, its level of intake and the diet in which the feed is fed
  - In situ and in vitro digestibility are affected methodology

# Objectives

- Discuss the factors that affect the in vitro and in situ measurement of NDFD
- Indicate the magnitude of variation in NDFD
- Discuss approaches to minimize variation in NDFD within and among laboratories

# NDFD Determination Basic Steps

- Material preparation
- Test sample selection
- Inoculum preparation
- Buffer
- Media supplementation
- Fermentation
- Residue collection

# Factors Affecting IV Digestibility

- Test sample preparation
  - Drying – less than 60C to minimize heat damaged protein and artifact lignin
  - Grinding recommendations vary
    - 8-mm screen Wiley cutter mill
      - Maximizes detection of physical effects
    - 2-mm screen Wiley cutter mill
      - Used for porous bag methods to minimize particle loss
      - Concentrates (1.5 to 2.5 mm), forages (1.5 to 5 mm)
    - 1-mm screen Wiley cutter mill
      - Most commonly used to detect digestibility differences
    - 1-mm screen, cyclone mill
      - Rarely, if ever, used for in vitro

# Effect of Wiley Grind Size on Corn Silage 24h IV Digestion

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<b>Size</b>	<b>IVDMTD</b>	<b>SD</b>	<b>IVNDFD</b>	<b>SD</b>
<b>Whole</b>	<b>73.2</b>	<b>5.69</b>	<b>37.6</b>	<b>9.27</b>
<b>4-mm screen</b>	<b>76.7</b>	<b>3.79</b>	<b>44.9</b>	<b>5.50</b>
<b>1-mm screen</b>	<b>77.4</b>	<b>3.96</b>	<b>48.7</b>	<b>5.33</b>

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Mertens and Ferreira (2000)

# Material Grind Size

- McLeod and Minson (1969) – Grasses Christy mill
  - 0.40 mm-screen = 54.3% T&T IVDMD 48h
  - 1.00 mm-screen = 52.4% T&T IVDMD 48h
  - 1.96 mm-screen = 49.7% T&T IVDMD 48h
- Alexander (1969) Christy mill
  - 0.60 mm = 53.8% ( $\pm 0.35$ ) T&T IVDMD 48h
  - 1.60 mm = 50.3% ( $\pm 0.70$ ) T&T IVDMD 48h
  - 2.45 mm = 50.1% ( $\pm 0.71$ ) T&T IVDMD 48h

# Material Grind Size

- Saldivar et al. (1982)
  - 0.5 UD = 52.5% T&T IVOMD 48h
  - 0.5 W = 52.3% T&T IVOMD 48h
  - 1.0UD = 50.3% T&T IVOMD 48h
  - 1.0 W = 47.1% T&T IVOMD 48h

# Factors Affecting In Vitro Digestibility

- **Sample amount**
  - Smaller amounts typically increase variation
  - **Flask/tube method**
    - Ratio of sample amount to buffer and inoculum
    - Typically .5 g per 40 ml buffer & 10 ml inoculum
  - **Bag method**
    - Ratio of sample amount to buffer and inoculum
    - Ratio of sample amount to bag surface area
    - Typically recommend 10 to 20 mg/cm<sup>2</sup>

# Test Sample Amount

- McLeod and Minson (1969) - Grasses
  - 0.5g = 58.0% T&T IVDMD 48h ( $\pm 1.1$ )
  - 0.6g = 57.2% T&T IVDMD 48h
  - 0.7g = 56.4% T&T IVDMD 48h
  - 0.8g = 56.1% T&T IVDMD 48h
  - 0.9g = 55.3% T&T IVDMD 48h
  - 1.0g = 55.0% T&T IVDMD 48h ( $\pm 0.5$ )

# Factors Affecting In Vitro Digestibility

- **Fermentation Vessel**
  - Flasks versus tubes
    - Changes surface area of submerged material
    - Changes side-wall contact
  - **Bag characteristics**
    - Size and area
      - 5X5 cm = 50 cm<sup>2</sup>
    - Type
      - Filter bag (F57)
      - Dacron bags
    - Pore size
      - 50 μm (range from 20 to 60 μm)

# Fermentation Vessel

- Sayre and Van Soest (1972)
  - Erlenmeyer flasks = 75.6% IVDMTD
  - Centrifuge tubes = 72.3% IVDMTD
  - Screwcap vials = 73.3% IVDMTD
- Robertson et al. (per. comm.)
  - 25 mm tubes = 52.3% IVNDFD
  - 32 mm tubes = 54.4% IVNDFD
  - Erlenmeyer flasks = 56.8% IVNDFD
- Grant and Mertens (1992)
  - 50 mL tubes = 66.3% IVNDFD
  - 125mL flasks = 67.8% IVNDFD

# Factors Affecting In Vitro Digestibility

- Buffer used to maintain pH during fermentation
  - McDougall's artificial saliva
  - Ohio buffer
  - Kansas buffer
  - Van Soest buffer

# Factors Affecting In Vitro Digestibility

- **Supplementation of media**
  - Trace minerals
  - Ammonia and amino acids
  - Branched-chain fatty acids
- **Reduction and anaerobicity**
  - Use of sulfide and cysteine
    - Reduced lag time (Grant and Mertens, 1992)
  - Use of indicator (resazurin)
  - CO<sub>2</sub> saturation of media and purging of vessels

# Flushing Vessels with CO<sub>2</sub>

- **Minson and McLeod (1972)**
  - Flushing gave no benefit for T&T IVDMD
    - 57.1% with versus 57.5% without
- **Alexander (1969)**
  - CO<sub>2</sub> buffer+CO<sub>2</sub> flush = 61.0% IVOMD 48h
  - CO<sub>2</sub> buffer+No flush = 59.4% IVOMD 48h
  - No buffer+No flush = 57.8% IVOMD 48h

# Flushing Vessels with CO<sub>2</sub>

- Robertson et al. (per. comm.)
  - Cont. manifold = 56.5% IVNDFD 48h
  - Bunsen valves = 52.4% IVNDFD 48h
- Grant and Mertens (1992)
  - Cont. manifold = 69.6% IVNDFD 48h
  - Purge + Bunsen = 58.4% IVNDFD 48h

# Factors Affecting In Vitro Digestibility

- **Sample wetting/submerging**
  - Floating material is a problem
    - Related to trapped gas and hydrophobicity
    - May interaction with vessel type
  - **Solutions**
    - Wet with a small amount of buffer
    - Submerge by evacuation
    - Swirling/mixing of vessels during fermentation
- **Clumping a material in bags**

# Test Sample Wetting

- Minson and McLeod (1972) used evacuation to submerge particles
  - IVDMD = 53.2% without versus 55.2% with

# Factors Affecting In Vitro Digestibility

- Inoculum Preparation

- Donor

- Single versus composite donors
    - Diet – Intake level
    - Feed restriction prior to obtaining contents
      - Fasting beyond 16 hr is detrimental (Ayers, 1991)

- Characteristics

- pH
    - Optical density

# Inoculum Preparation

- Ayres (1991)
  - Sheep W19 52.6% IVOMD
  - Sheep W34 51.2% IVOMD
  - Sheep W26 46.6% IVOMD
  - Sheep W31 45.1% IVOMD
  - Composite 51.6% IVOMD
- Mertens, Weimer & Waghorn (unpubl)
  - Composite performed better than individual donors

# Strained Ruminant Fluid pH/OD

- McLeod and Minson (1969) - Grasses
  - pH 6.1 = 58.8% T&T IVDMD 48h
  - pH 6.7 = 59.2% T&T IVDMD 48h
  - pH 7.2 = 62.5% T&T IVDMD 48h
- Mertens and Ferriera (unpubl)
  - IVNDFD reduced below an OD threshold

# Factors Affecting In Vitro Digestibility

- Inoculum Preparation
  - Strained rumen fluid versus solids extraction
    - Particle associated microbes
  - Time from collection to inoculation
  - Amount of inoculum

# Inoculum Preparation

- Craig et al. (1984)
  - Particle-associated microbes collected by washing strained ruminal solid (+PM)
  - Solids were blended with ruminal fluid (B)
  - SRF = 46.3% IVNDFD 48h
  - SRF+PM = 48.6% IVNDFD 48h
  - SRF(B) = 46.1% IVNDFD 48h
  - SRF+PM4°C = 45.8% IVNDFD 48h

# Inoculum Preparation Delay

- **Alexander (1969)**
  - Normal (15min) 68.4% T&T IVDMD 48h
  - 1h delay 38.5 °C 62.3% T&T IVDMD 48h
  - 1h delay cooled 58.3% T&T IVDMD 48h
- **Mertens (1973)**
  - Delay beyond 20 min (cow to inoculation) increased lag time

# Strained Rumen Fluid to Buffer Ratio

- McLeod and Minson (1969) – Grasses
  - 25:25 = 52.9% T&T IVDMD 48h
  - 15:35 = 51.2% T&T IVDMD 48h
  - 10:40 = 48.5% T&T IVDMD 48h
  - 5:45 = 43.9% T&T IVDMD 48h
  - 2.5:47.5 = 43.9% T&T IVDMD 48h
- Weimer (per. comm.)
  - IV digestion reduced below 10 mL SRF

# Factors Affecting In Vitro Digestibility

- Incubation temperature
  - Recommended varies from 38-39.5 °C
  - Gas pressure measurements were extremely sensitive (Mertens and Weimer)
    - 10% reduction per 1 °C difference from 39 °C

# Incubator Temperature

- Alexander (1969)

– 35.5 °C	= 56.4% IVOMD 48h
– 38.5 °C	= 58.7% IVOMD 48h
– 42.0 °C	= 61.0% IVOMD 48h

- Minson and McLeod (1972)

– 35.0 °C	= 54.4% IVOMD 48h
– 37.0 °C	= 58.4% IVOMD 48h
– 39.0 °C (min SD)	= 58.9% IVOMD 48h
– 41.0 °C	= 59.7% IVOMD 48h
– 43.0 °C	= 58.4% IVOMD 48h

# Factors Affecting In Vitro Digestibility

- Adjustment using standards
  - Traditionally used in vitro versus in vivo calibration curves
    - Required 4 to 5 calibration samples per run
    - Variable effectiveness
  - Use standards to normalize or correct individual results
  - Use standards to determine validity of the entire run without correction

# Adjustment of IV Digestibility Using Standards

- Adjustment using standards
  - Alexander (1969)
    - Scaling for std mean was ineffectual
    - Correcting using 4 ref std reduced single result SD from 1.27 to 0.89
  - Ayers (1991)
    - No adjustment if standards with 95% confidence level
    - Adjust by mean deviation, if the deviations of 4 standards are consistently different
    - Re-run if standards outside the 95% CI and are inconsistent
  - Mertens
    - Using standards as covariate rarely improves statistical analysis

# Factors Affecting In Vitro Digestibility

- Time of fermentation
  - T&T 48h IVDMD consistently related to in vivo digestibility measured at maintenance levels of intake (Feed Evaluation Protocol)
  - Allen et al indicate that producing dairy cows have a fiber retention time of 30 to 36 h
  - Some have suggested that 24h IV fermentations may be a better indication of dairy cow performance

# In Vitro Fermentation Time versus In Vivo Retention Time

- In vivo Retention Time DOES NOT equal in vitro fermentation time
  - i.e., digestion at 30 hr retention time DOES NOT equal digestion at 30 hr fermentation time
    - $1/k_p = \text{retention time} \neq \text{fermentation time}$
  - In vivo digestion =  $k_d / (k_d + k_p)$
  - In vitro digestion =  $1 - DM \cdot \exp(-k \cdot t)$

# In Vitro Variation

- Alexander (1969)
  - 1-stage rumen fluid
    - Between run      SD = 0.99
    - Within run        SD = 0.73
  - 2-stage rumen fluid + acid pepsin (T&T)
    - Between run      SD = 0.63
    - Within run        SD = 0.38

# In Vitro Variation

<b>Reference</b>	<b>Within run</b>	<b>Among run</b>
<b>Alexander (1969)</b>	<b>.39</b>	<b>.66</b>
<b>Tilley &amp; Terry (1963)</b>	<b>.61</b>	<b>1.90</b>
<b>Dent (1963)</b>		<b>1.50</b>
<b>Minson and McLeod (1972)</b>	<b>.94</b>	<b>2.24</b>
<b>Martin and Barnes (1969) A</b>	<b>.83</b>	
<b>Martin and Barnes (1969) B</b>	<b>.50</b>	
<b>Barnes (1967) 5-lab average</b>	<b>2.80</b>	<b>2.35</b>
<b>Ayers (1991)</b>		<b>1.18</b>

# In Vitro Variation

- IVNDFD is more variable than IVDMTD or IVDMMD
  - IV undigested NDF (uNDF) has a variance
  - NDF determination has a variance
  - IVNDFD is the quotient of two variables
    - $IVNDFD = 100 * (NDF - uNDF) / NDF$
  - Mathematical consequence of dividing mean and SD by a fraction
    - Mean = 50 and SD = 5, if all measurements are divided by .5 then Mean = 100 and SD = 10

# NDFD Variation – Statistics 101

- **Summation of errors**
  - SD of determining NDFD using in vitro method =  $\pm 4.0$
  - SD of predicting IVNDFD using NIRS =  $\pm 3.0$
  - Total SD of estimating NDFD using NIRS
    - = square root ( $IV\_SD^2 + NIRS\_SD^2$ ) =  $\pm 5.0$
- **Outlier population**
  - $\pm 1$  SD = 31.7% of estimates outside  $\pm 5$
  - $\pm 2$  SD = 4.6% of estimates outside  $\pm 10$
  - $\pm 3$  SD = 0.26% of estimates outside  $\pm 15$

# In Vitro Digestibility – Final Caution

- IVDMD DOES NOT EQUAL in vivo DMD, especially at production levels of performance
- Improvement in IVDMD and IVNDFD of bmr corn does not translate into improved dairy cow digestibility
  - Instead performance is increased due to increased intake
  - Not certain this is a universal response, but should indicate caution in using in vitro data